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New species of Arctiinae east of the Canal Zone, Panama (Lepidoptera, Erebidae, Arctiinae, Arctiini)

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description;
Canal Zone;
Panama.

Abstract. – A study of a fresh material received from Panama, East of the Canal Zone, has revealed five new species which are then described: *Robinsonia xanthopoda* **sp**. **n**., *Syntomostola harrisoni* **sp**. **n**., *Pseudosphex lindae* **sp**. **n**., *Pseudalus panamensis* **sp**. **n**., and *Trichromia croceicervix* **sp**. **n**. Moreover new highlights on poorly known species allowed new recombinations, synonymies or new status for several species in the genera *Pryteria* Möschler, 1883, *Marecidia* Schaus, 1901 or *Sphecosoma* Butler, 1876. Finally for the first time, male genitalia are provided for some species in the genera *Rhipha* Walker, 1854 or *Hyperthaema* Schaus, 1901.

Laguerre M., 2025. – New species of Arctiinae east of the Canal Zone, Panama (Lepidoptera, Erebidae, Arctiinae, Arctiini). *Faunitaxys*, 13(12): 1 – 17.

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Introduction

Since 2021 I have received very interesting material from Panama thanks to Linda & Jerry Harrison. The material was collected east of the Canal Zone, 23 km NNE of Panama City at an altitude of 700 m, in the close vicinity of Cerro Azul and Cerro Jefe (9.2030° N 79.4150° W). The fauna in this area seems largely different of the classical fauna found in the Chiriqui area with a high endemicity. This area makes a link with the Pacific Coast in Colombia and western Ecuador by the way of the Darien Province. Several new species of Arctiinae were found but also species collected bring new highlights on several poorly known entities. The results of this study are provided below with descriptions of five new species and discussions on several species insufficiently or hardly characterized.

Materials and Methods

This paragraph has been treated in details in several previous papers, some of them being easily available (Laguerre, 2023 and Laguerre *et al.,* 2023), and it will not be repeated there.

Repository abbreviations

LMK	Landesmuseum Kärtern, Klagenfurt, Austria.
MNHN	Muséum National d'Histoire Naturelle, Paris, France.
MNHU	Museum für Naturkunde, Humboldt-Universität, Berlin, Allemagne.
MUSM	Museo de Historia Natural, Univ. Mayor de San Marcos, Lima, Peru.
NHMUK	Natural History Museum, London, England, United Kingdom.
PMJ	Phyletisches Museum, Jena, Germany.
USNM	United States National Museum, Smithsonian Institution, Washington D. C., U. S. A.
MLC	Personal collection of Michel Laguerre, 33850 Léognan, France.
VDC	Personal collection of Vyacheslav Doroschkin, Tcheliabinsk, Russia.

Abbreviations used

FWL = ForeWing Length (from base to apex)
WS = WingSpan

Results

Descriptions of new species

One of the first images received was a male *Robinsonia*, the external appearance was similar to species of the group of *R. praphoea* Dognin, 1906: wings entirely silky white and a thin costal black border, but the legs annealed with orange were surprising and unique (see picture below). Clearly the specimen was a new species described below under the name *Robinsonia xanthopoda* **sp**. **n**.

***Robinsonia xanthopoda* sp. n.**

ZooBank: https://zoobank.org/1FF8A590-F528-4E29-BC15-F44BF6B53B11

Holotype, ♂, PANAMA, Panama, Los Altos de Cerro Azul, 14-XI-2023, 700 m, 9.2030° N 79.4150° W, L. & J. Harrison *leg*., Will be deposited in MNHN.

Paratypes (**2** ♂, **4** ♀), same locality as holotype, in MLC.

- **2** ♂, 13-VII-2021, one is dissected Gen. ML3828 (light-blue manuscript label);
- **2** ♀, 13-XII-2021 (dissected ML3882) and 23-VII-2021;
- **2** ♀ 14-XI-2023, D. Herbin *leg*., (one is dissected ML3881).

Diagnosis. – An entirely silky white species, with a thin black line on forewing costa and an orange vertex. Legs annealed with black and orange, forecoxae orange in males.

Male (Fig. 2)

FWL = 14 - 15 mm. Antennae black, bipectinate. Head with frons and vertex bright orange. Palpi orange. Collar white. Tegulae, patagia and thorax white. Abdomen above covered with dirty white hairs. Forewings entirely silky white

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Fig. **1**. Living picture of a ♂ of *Robinsonia xanthopoda* **sp**. **n**. in Panama (L. & J. Harrison courtesy).

with a thin black border along costa. In some specimens a trace of grey shading on the veins terminus below apex. Hindwings entirely silky white. Below forecoxae bright-orange. Fore- and mid-legs with tarsi dark grey, fore- and mid-femurs dark grey annealed with orange, hind-legs dirty white. Abdomen below dirty white. Underside of wings similar to upperside, the forewing costa border wider and the grey shading on the veins below apex more pronounced.

Genitalia male (Fig. 3)

Uncus moderately wide, enlarged at base and then flattened and curved downward. Valvae not reaching the tip of uncus, wider at base then spoon like in the last half with a rounded tip. A translucent and rounded sacculus covered with thin hairs. Juxta conical. Vinculum flat slightly concave without saccus. Aedeagus long, narrow and cylindrical with a pronounced caecum penis. Vesica smooth, bilobed: one short lobe and one very long tube, narrow and cylindrical with a short diverticule near base bearing four teeth at the extremity.

Female (Fig. 4).

FWL = 16-17 mm. Similar to male, slightly larger, with forewings less elongated. Head with frons and vertex yellow. Palpi beige. Forecoxae light yellow. The black costa line is thinner, especially below. The short dark streaks on the forewing preapical termen below are absent. The legs are less contrasted.

Genitalia female (Fig. 5)

Papillae anales subrectangular, slightly concave in middle. Apophyses anteriores long and thin, apophyses posteriores shorter and wider. Antrum straight, ductus bursae as a flat ribbon, folded in the middle. Bursa copulatrix spherical and wrinkled with two symmetrical signa like a rounded corrugated depression. The bursa copulatrix is linked to an oval accessory bursa by the way of a sclerotized cylindrical tube bent at 90° of the ductus bursae and turned on the right in ventral view.

Fig. **2**. Holotype ♂ of *Robinsonia xanthopoda* **sp**. **n**. upperside on left, underside on right, labels not to scale.

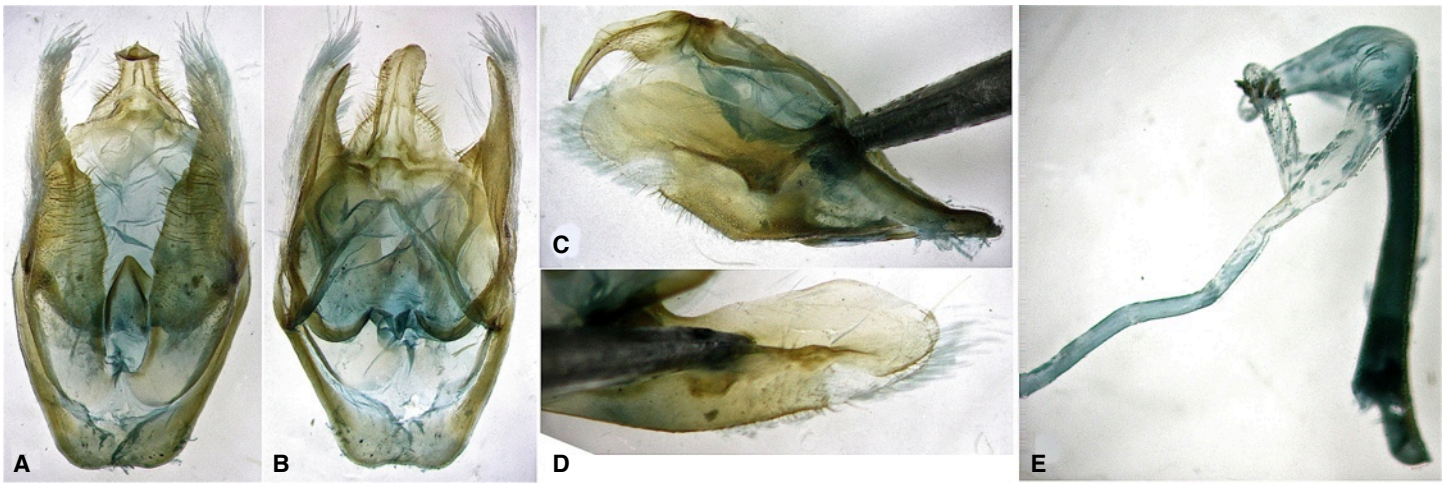

Fig. **3**. Genitalia of ♂ paratype of *Robinsonia xanthopoda* **sp**. **n**. from Panama (ML3828). **A**. Ventral view. **B**. Dorsal view. **C**. Lateral view. **D**. Valva inside view. **E**. Aedeagus with everted vesica.

Fig. **4**. Paratype ♀ of *Robinsonia xanthopoda* **sp**. **n**. upperside on left, underside on right, labels not to scale.

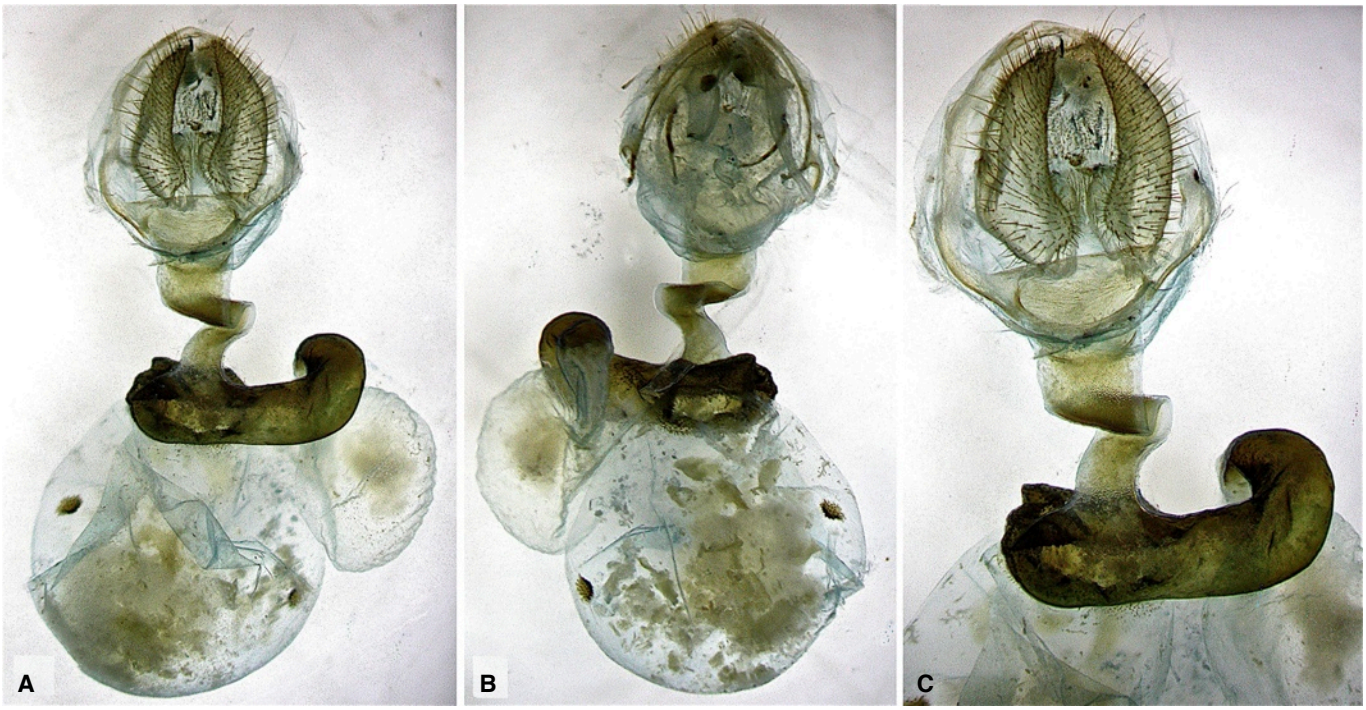

Fig. **5**. Genitalia of ♀ paratype of *Robinsonia xanthopoda* **sp**. **n**. from Panama (ML3881). **A**. Ventral view. **B**. Dorsal view. **C**. Detail view.

Etymology. – The name *xanthopoda* derives from the Ancient Greek xanthos (ξανθος) which means yellow and poda (ποδα) which means feet or legs, by reference to the legs characteristic color.

Early stages. – Unknown.

Distribution. – Presently only known from Panama East of the Canal Zone at low- to mid-altitudes.

Syntomostola harrisoni **sp**. **n**.

ZooBank: https://zoobank.org/3F94B251-C8F8-4D99-AD21-F85170B7915F

The first contact with this species occurred in 2009 during a visit to the Smithsonian Institution in Washington. A single male specimen was found in a drawer of specimens awaiting study. Since that, no other specimen was found until a first picture received from Panama (see Fig. 6). Finally several male and female specimens were received and this species is in fact a *Syntomostola* close to *S. xanthosoma* Dognin, 1912 but clearly undescribed. The species will be described as new below under the name *Syntomostola harrisoni* **sp**. **n**.

Holotype, ♂, PANAMA, Panama, Los Altos de Cerro Azul, 14-XI-2023, 700 m, 9.2030° N 79.4150° W, L. & J. Harrison *leg*., dissected Gen. ML3959 (light-blue manuscript name). Will be deposited in MNHN.

Paratypes (**4** ♀), same data as holotype but 08-IX-2020, 05-I-2021, 24-II-2021 and 27-VI-2021. In MLC.

Diagnosis. – The colour pattern is characteristic and unique. The forewings are bright yellow to orange-yellow basally and then pure black up to the apex. There is a large transparent postmedian transverse band from costa near to tornus, with black veins and a triangular transparent area in cell. The whole body is bright yellow to orange-yellow except the head which is black. The legs are black with yellow forecoxae.

Male (Fig. 7)

FWL = 14 mm. Antennae thin and long, black and ciliate. Head and palpi entirely black. Proboscis brown. Collar, patagia, tegulae, thorax and abdomen entirely yellow or orange-yellow. Forewings elongated, yellow or orange-yellow basally, then black up to the apex. A large transverse band from costa to near tornus, transparent crossed by black veins. A triangular transparent area on cell crossed by a black vein. Costa black except base which is yellow. Hindwings semi-oval, elongated vertically, the basal two-thirds yellow, the outer third black with an irregular internal border. Underside identical to upperside. The forecoxae hairy and yellow, the legs black. Abdomen underside yellow.

Genitalia male (Fig. 8)

Genitalia slightly asymmetrical, the majority of the genital capsule is covered with thin and short hairs. Uncus in frontal view as a very

narrow oval, with a dorsal process shaped as a short blade. On the tegumen, above uncus and on each side a digitate process pointing frontally with a rounded tip. Valvae outreaching the uncus and processes, narrow with a rounded extremity. Near base a strong sclerotized torn. Juxta bulbous. Vinculum evenly rounded with a very narrow pointed saccus turn to left in ventral view. Aedeagus cylindrical, bent at 90° in the last half. A short caecum penis is present. Vesica with a unique tubular and scobinate lobe turned at 180° and terminated by a long and narrow tube.

Female (Fig. 9)

FWL = 14-15 mm. Similar to male with rounder forewings. Hindwings are not elongated vertically. The forewing costa is more yellow basally. On the hindwings the black border is wider and the yellow colour is restrained to the base of the wings.

Etymology. – By reference to Linda and Jerry Harrison who provided the totality of the type series.

Early stages. – Unknown.

Distribution. – Presently known only from Panama at low to mid-altitudes.

Except the present species, the genus *Syntomostola* contains two more species: *S. xanthosoma* Dognin, 1912 described from Colombia, Medina (holotype ♂ in USNM) and *S. semiflava* Dognin, 1923 described from High Amazon, Brazil (holotype ♂ in USNM). The two types were dissected by Watson in 1971 (plate 138) and the two dissections appear similar but in the absence of any specimen of *S. semiflava* we maintain the status quo. A specimen from Cundinamarca, Colombia (Amazonian foothills at 500 m) was sequenced and its sequence (reference MILA2332) aligns with several species of *Trichromia* or *Haemanota*, so confirming it belongs to the Phaegopterina subtribe. It is interesting to note that Draudt in Seitz described in 1915 a female of the same species in the Ctenuchina subtribe under the name *Xantharete stellans*. The type locality, Colombia, Rio Negro is coherent with the sequenced specimen. So we will have *Syntomostola xanthosoma* Dognin, 1912 = *Xantharete stellans* Draudt, 1915, **syn**. **nov**. Moreover as *Xantharete* is a monospecific genus we will have also *Syntomostola* Dognin, 1912 = *Xantharete* Draudt, 1915 **syn**. **nov**.

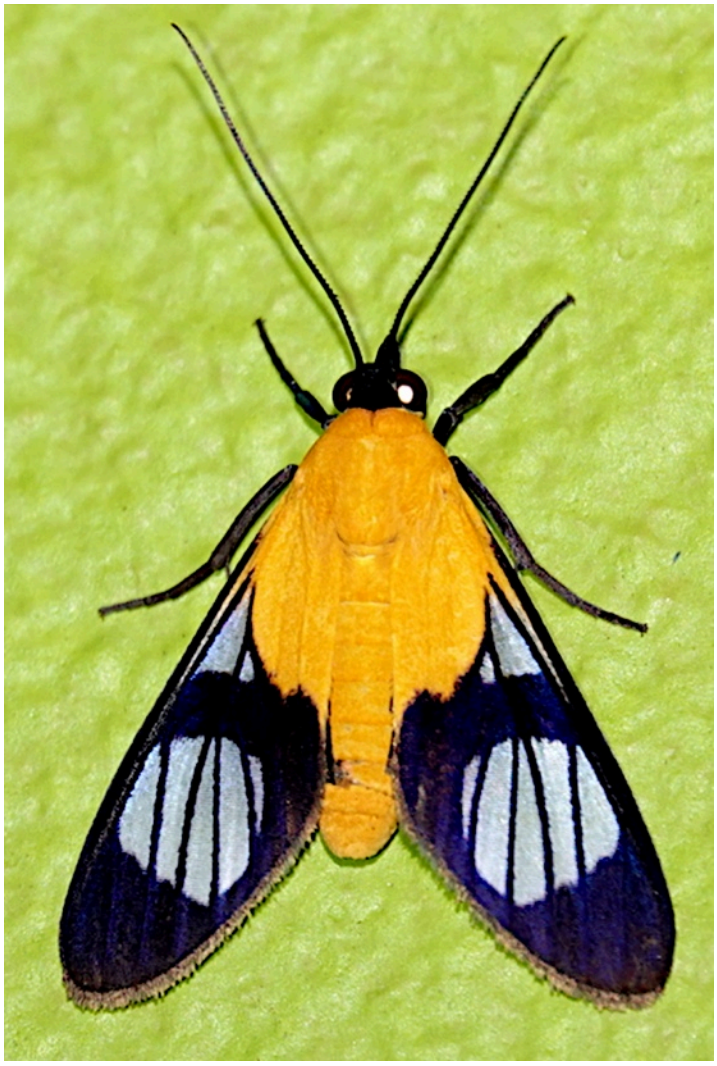

Fig. **6**. Living picture of *Syntomostola harrisoni* **sp**. **n**. in Panama (L. & J. Harrison courtesy).

Fig. **7**. Holotype ♂ of *Syntomostola harrisoni* **sp**. **n**. upperside on left, underside on right, labels not to scale.

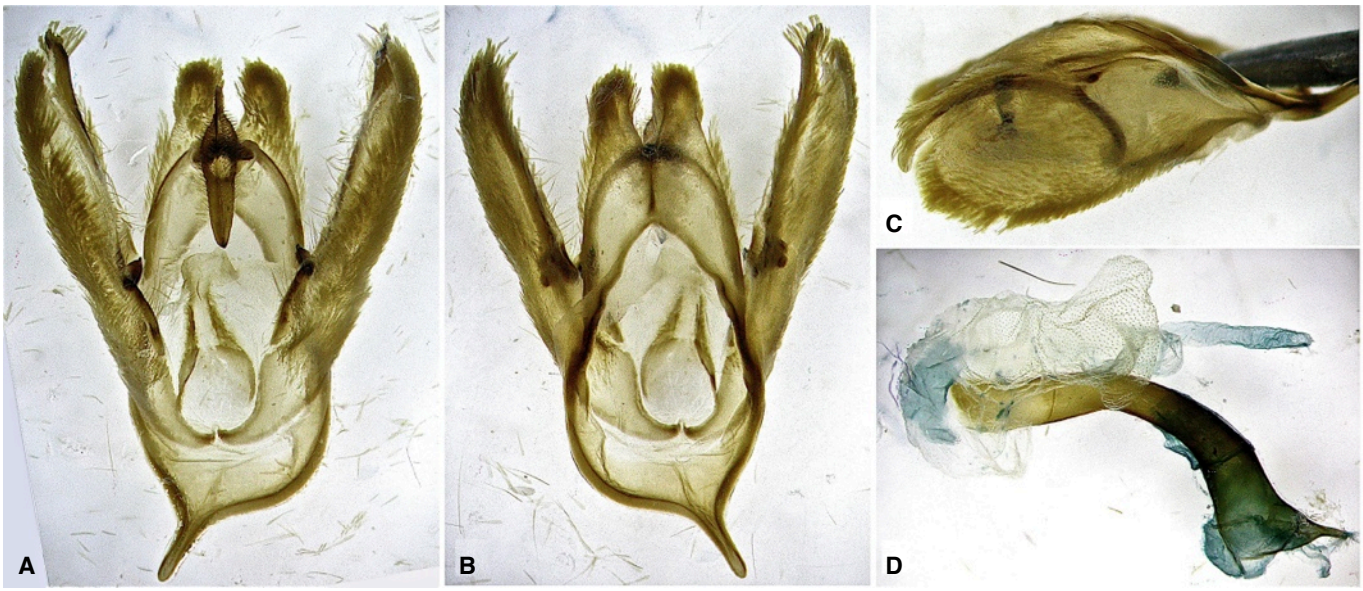

Fig. **8**. Genitalia of holotype ♂ of *Syntomostola harrisoni* **sp**. **n**. from Panama (Gen. ML3959). **A**. Ventral view. **B**. Dorsal view. **C**. Lateral view. **D**. Aedeagus with everted vesica.

Fig. **9**. Paratype ♀ of *Syntomostola harrisoni* **sp**. **n**. upperside on left, underside on right, labels not to scale.

***Pseudosphex lindae* sp. n.**

ZooBank: https://zoobank.org/5FAB1EF3-24F6-4CC2-848C-2E7F64A44D13

In the material received we found several specimens of a *Pseudosphex* species very similar to *P. fulvisphex* (Druce, 1898) but some differences in the habitus clearly justify a careful study (see below Fig. 10).
After dissection, it appears that, even if the external appearance is very close to *Pseudosphex fulvisphex*, the male genitalia surprisingly display a pattern similar to those of *Pseudosphex aequalis* (Walker, [1865]) despite different habitus. Nevertheless, the genitalia are divergent enough to consider this entity as new and it will be described below under the name *Pseudosphex lindae* **sp**. **nov**.

Holotype, ♂, PANAMA, Panama, Los Altos de Cerro Azul, 27-XII-2020, 700 m, 9.2030° N 79.4150° W, L. & J. Harrison *leg*., dissected Gen. ML3856 (light-blue manuscript label). Will be deposited in MNHN.

Paratypes (**3** ♀), same data as holotype but 05-V-2023 (dissected ML3829, not shown), 28-III-2023 and 17-VIII-2023. In MLC.

Diagnosis. – Species wasp mimic, very close to *Pseudosphex fulvisphex*. Wings entirely transparent with black veins. Costa orange and a thin black border enlarged at apex on forewings only.

Male (Fig. 11)

FWL = 12 mm. Antennae black with long ciliae in the basal two-thirds, scape black with an orange dot. Head, frons and vertex yellow. A black transverse line before collar. Patagia orange with a central black bar. Tegulae orange with a central black band. Thorax dark orange with a thin median black line and two transverse black bands, one just after the patagia, the second in middle position. Abdomen upperside orange, the two first segments pale yellow with a thin central black line. A black spot on each side of the first segment. The remaining segments orange with a transverse black band just after the yellow segments. Forewings entirely transparent with black veins. A thin black border enlarged at apex. Costa orange from base to just before apex. Internal border orange fading toward tornus. Hindwings entirely transparent with very thin black border and orange costa. Underside: palpi, forecoxae legs and abdomen underside orange.

Genitalia male (Fig. 12)

Uncus wide at base, evenly bent ventrally, finishing with a sclerotized pointed tip. Valvae subrectangular with a pointed triangular extremity. Just above this extremity a very long and narrow arched process largely outreaching the tip of uncus. This process displays a ventral slightly crenulated carina. Presence of a slightly sclerotized subscaphium. Juxta shaped as a tongue reaching the base of uncus. Vinculum curved with a long saccus with a rounded tip. Aedeagus long, cylindrical and narrow, smoothly bent at 90° in the last half. Vesica smooth with a long unique lobe bent at 180° and terminated with a long and narrow tube.

Female (Fig. 13)

FWL = 13-14 mm. Similar to male. Slightly larger and the black border on wings is wider. The antennae have very short ciliae.

Etymology. – By reference to Linda Harrison who, with husband Jerry, provided almost the totality of the material at the base of this study.

Early stages. – Unknown.

Distribution. – Presently only known from Panama around the Canal Zone.

Fig. **10**. Living picture of a pair of *Pseudosphex lindae* **sp**. **n**. in Panama (L. & J. Harrison courtesy), ♂ on the left, ♀ on the right.

Fig. **11**. Holotype ♂ of *Pseudosphex lindae* **sp**. **n**. upperside on left, underside on right.

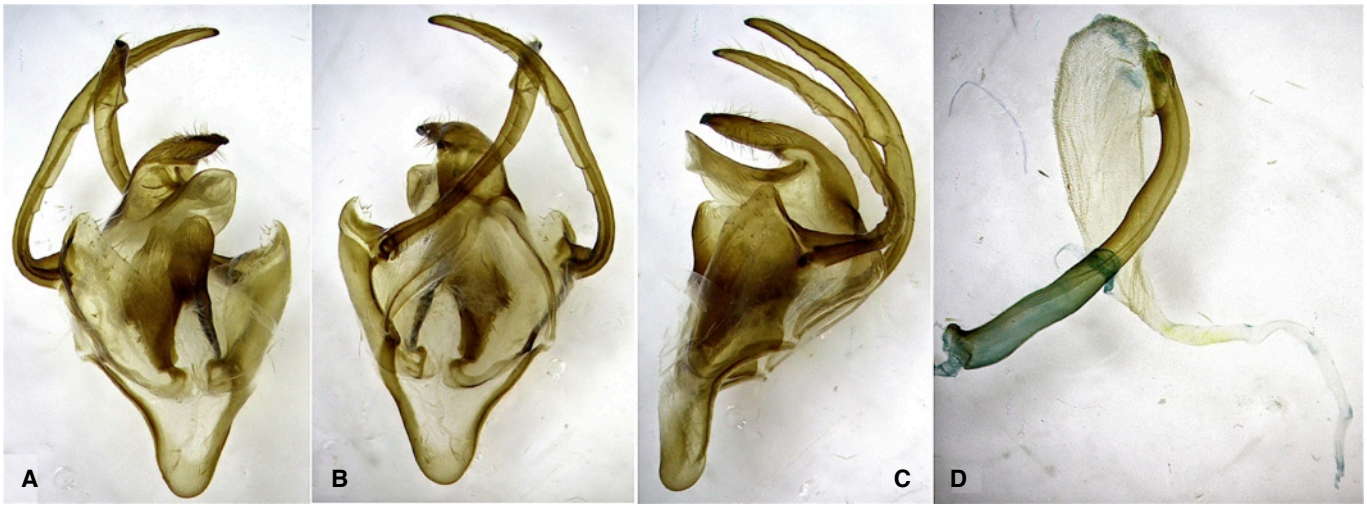

Fig. **12**. genitalia of holotype ♂ of *Pseudosphex lindae* **sp**. **n**. from Panama (ML3856). **A**. Ventral view. **B**. Dorsal view. **C**. Lateral view. **D**. Aedeagus with everted vesica.

Fig. **13**. Paratype ♀ of *Pseudosphex lindae* **sp**. **n**. upperside on left, underside on right.

***Pseudalus panamensis* sp**. **n**.

ZooBank: https://zoobank.org/FD45E11F-4AB3-4FEC-8EEB-55D0412A2545

In the material received, there was a series of a small yellow *Pseudalus* of the *leos/affinis* group. The genus *Pseudalus* Schaus, 1896, contains now eight species and one subspecies and is largely neglected presently. The specimens collected in Panama have an external appearance similar to the type of *P. affinis* Rothschild, 1933 (NHMUK) and a specimen collected in the Cerro Hoya National Park was sequenced. Only 17 *Pseudalus* sequences were found in BOLD and the resulting Maximum Likelihood tree is shown below (Fig. 14). It then appears that the *Pseudalus* from Panama aligns at 7.3 % from *Pseudalus leos occidentalis* Rothschild, 1909, from Peru (Cusco) and at 10.9 % from two specimens of *Pseudalus affinis*, from French Guiana. So clearly the Panama specimens represent a totally different species despite very similar habitus and they will be described as new below under the name *Pseudalus panamensis* **sp. nov.**

Holotype, ♂, PANAMA, Panama, Los Altos de Cerro Azul, 05-XII-2021, 700 m, 9.2030° N 79.4150° W, L. & J. Harrison *leg*., dissected Gen. ML3879 (light-blue manuscript label). Will be deposited in MNHN.

Paratypes (**2** ♂), same data as holotype but 04 and 05-XII-2021. In MLC.

Diagnosis. – A small yellow moth with two reddish-brown transverse lines on forewings, one weak and antemedian, the second postmedian not reaching exactly the apex and a tiny dot at the end of the cell.

Male (Fig. 15)

FWL = 11-12 mm. Antennae short and ciliate, dark yellow. Head, frons and vertex yellowish orange. Palpi brownish orange. Collar, patagia, tegulae, thorax and abdomen upperside yellowish. A vertical orange streak on thorax. Forewings yellow with two transverse reddish brown lines. The first one weak in antemedian position, the second postmedian, clear-cut, from middle of internal border to near apex. A tiny brownish-red dot at the end of cell. Hindwings plain yellow. Underside identical to upperside. The two transverse lines are indistinct, the first one generally absent, the second blurred and faded, the cell dot visible. Forecoxae, legs and abdomen underside brownish yellow.

Genitalia male (Fig. 16)

Extremely similar to the genitalia of *Pseudalus affinis* from French Guiana (see Fig. 17 below). Genital capsule very elongated vertically. Uncus large and hemispherical, slightly tightened at base, then narrow and compressed laterally, strongly hooked at tip. On the tegumen, above uncus and on each side, a short digitate process pointing frontally. Valvae just reaching the tip of uncus, subrectangular at base then narrowed and finishing as a blunt and thin blade. Juxta rhomboid. Vinculum evenly rounded with a triangular pointed saccus. Aedeagus narrow and cylindrical slightly curved toward extremity. A short caecum penis present. Vesica bilobed. A first lobe at 90°, short with a strongly scobinate and denticulate tip. A second lobe longer at 180° with a scobinate extremity bent toward the aedeagus. Just before this extremity a small digitate diverticule adorned with small teeth.

Etymology. – By reference to the locality of the type series.

Early stages. – Unknown.

Distribution. – Presently only known from Panama around the Canal Zone.

Fig. **14**. Maximum Likelihood tree of 17 *Pseudalus* sequences, bootstrap values in bold if > 70 %, branch lengths below lines if > 0.002.

Fig. **15**. Holotype ♂ of *Pseudalus panamensis* **sp**. **n**. upperside on left, underside on right.

Fig. **16**. Genitalia holotype ♂ of *Pseudalus panamensis* **sp**. **n**. from Panama (ML3879). **A**. Ventral view. **B**. Dorsal view. **C**. Lateral view. **D**. Aedeagus with everted vesica.

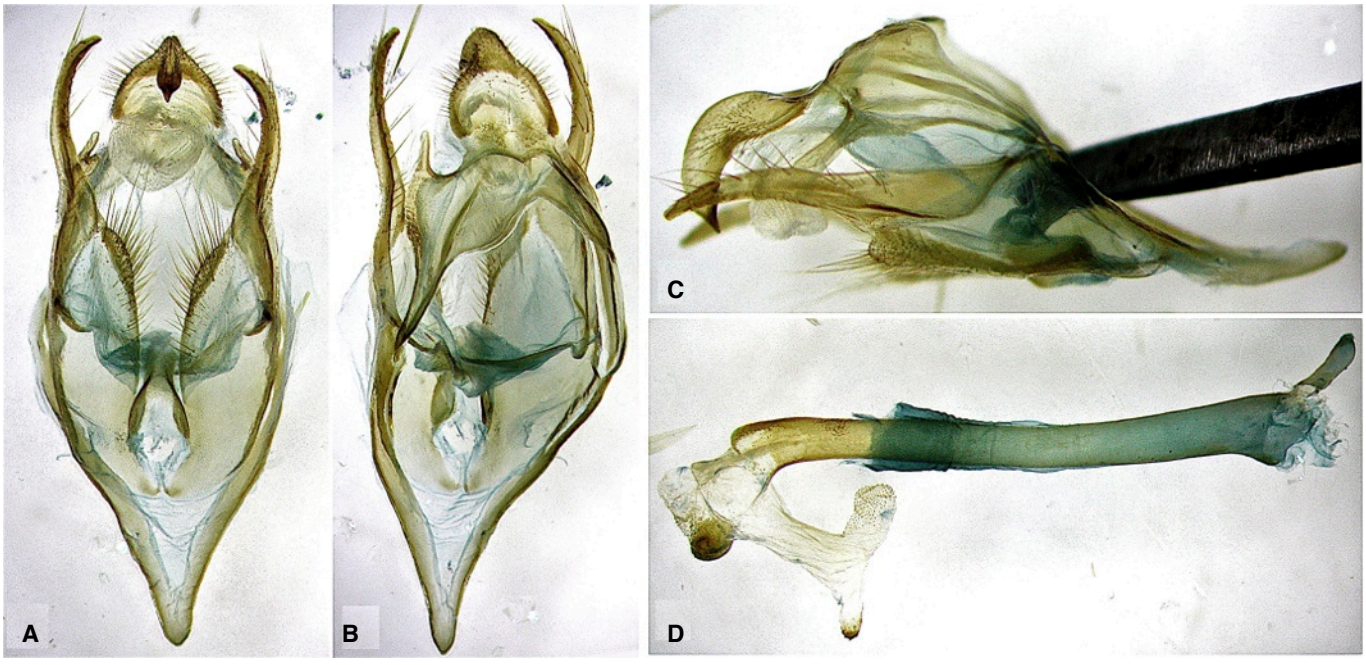

Fig. **17**. Genitalia ♂ of *Pseudalus affinis* from French Guiana (ML3680). **A**. Ventral view. **B**. Dorsal view. **C**. Lateral view. **D**. Aedeagus with everted vesica.

It is rather surprising to notice that the genus *Pseudalus* is presently classified within the sub-tribe Arctiina, whereas in a general tree of more than 15,000 Neotropical Arctiinae sequences, the genus is a close neighbour of the genus *Neidalia* Hampson, 1901, classified at the beginning of the sub-tribe Phaegopterina. The two genera align also close to *Hyperthaema* Schaus, 1901, *Rhodorhipha* Laguerre, 2018 or some *Zatrephes* Hübner, [1819]. So clearly *Pseudalus* is a member of the sub-tribe Phaegopterina, so we will have *Pseudalus* **comb. nov**. Phaegopterina. Moreover in this tree the genus *Neidalia* seems polyphyletic and is split into two clusters by the genus *Pseudalus*, but this problem is outside the scope of this paper and will be treated elsewhere.

The group of *Trichromia leucoplaga* (Hampson, 1905)

This group is problematic as, in BOLD, for specimens externally identical we have three clusters: one in Central America and two in French Guiana. There are also specimens from South-East Brazil but they were not sequenced. *T. leucoplaga* was described from Saint Laurent du Maroni in French Guiana but we have two sequences in French Guiana at a minimum distance of 7.1 % (see simplified tree Fig. 18 below) and as the type was neither dissected nor sequenced we don't know which of the two species present in French Guiana is the true *T. leucoplaga*.
Nevertheless the species from Central America which is respectively at 2.8 % and 6.7 % from the two species from French Guiana is surely new and will be described below under the name *Trichromia croceicervix* **sp**. **n**.

Trichromia croceicervix **sp**. **n**.

ZooBank: https://zoobank.org/435A5403-B72D-444F-930B-60FB01216572

Holotype, ♂, PANAMA, Panama, Los Altos de Cerro Azul, 03-I-2021, 700 m, 9.2030° N 79.4150° W, L. & J. Harrison *leg*., dissected Gen. ML3855 (ligh-blue manuscript label). Will be deposited in MNHN.

Paratypes. In MLC.
- PANAMA, **1** ♂, same data as holotype but 06-I-2021.
- NICARAGUA, **2** ♂, Granada, Volcan Mombacho, Cutirre, 20-I-2004, 687 m, J.M. Maes *leg*.
One is dissected Gen. ML767 (white manuscript label), the second is sequenced Process ID ARCTD086-11, BOLD ID MILA 1591 (yellow printed label).

Diagnosis. – A species identical to *Trichromia leucoplaga* and impossible to distinguish by the external appearance only.

Male (Fig. 19).

FWL = 15-16 mm. Almost impossible to distinguish from *Trichromia leucoplaga* from French Guiana. Nevertheless seems slightly larger and more robust: wingspan 32-33 mm against 29-30 mm for specimens from French Guiana. The vertex seems more orange, generally almost totally pale ochreous brown in *T. leucoplaga* except a thin orange band near collar. Underside the last third of the abdomen is washed with orange which is not the case in *T. leucoplaga*.

Genitalia male (Fig. 20)

Extremely homogeneous in the group and almost impossible to differentiate. Genital capsule subrectangular. Uncus long and narrow with a bifid extremity. On the tegumen, above uncus and on each side a strong sclerotized curved process pointing inward. The two processes cross slightly just above uncus. Their extremity is strongly narrowed at the end finishing as a long spine. The terminal part is denticulate frontally. Valvae

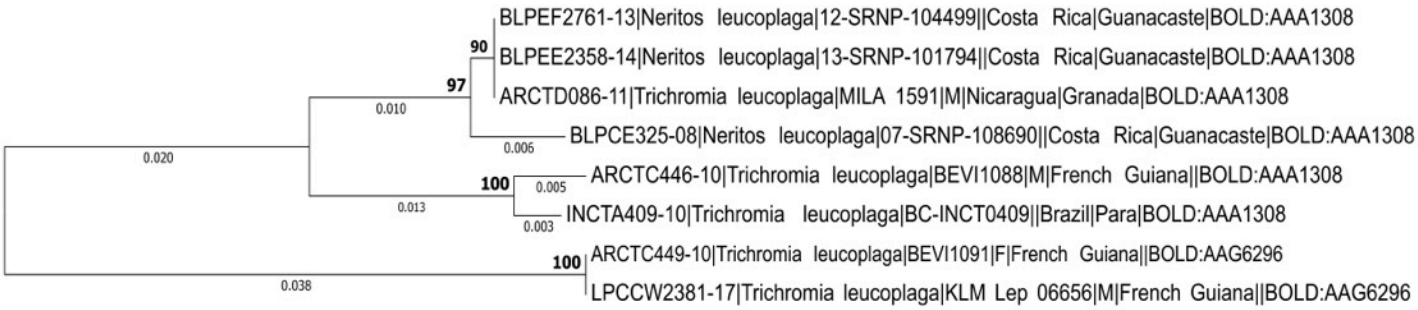

Fig. **18**. Maximum Likelihood simplified tree of *Trichromia leucoplaga* sequences, bootstrap values in bold if > 70 %, branch lengths below lines if > 0.002.

Fig. **19**. Holotype ♂ of *Trichromia croceicervix* **sp**. **n**. upperside on left, underside on right.

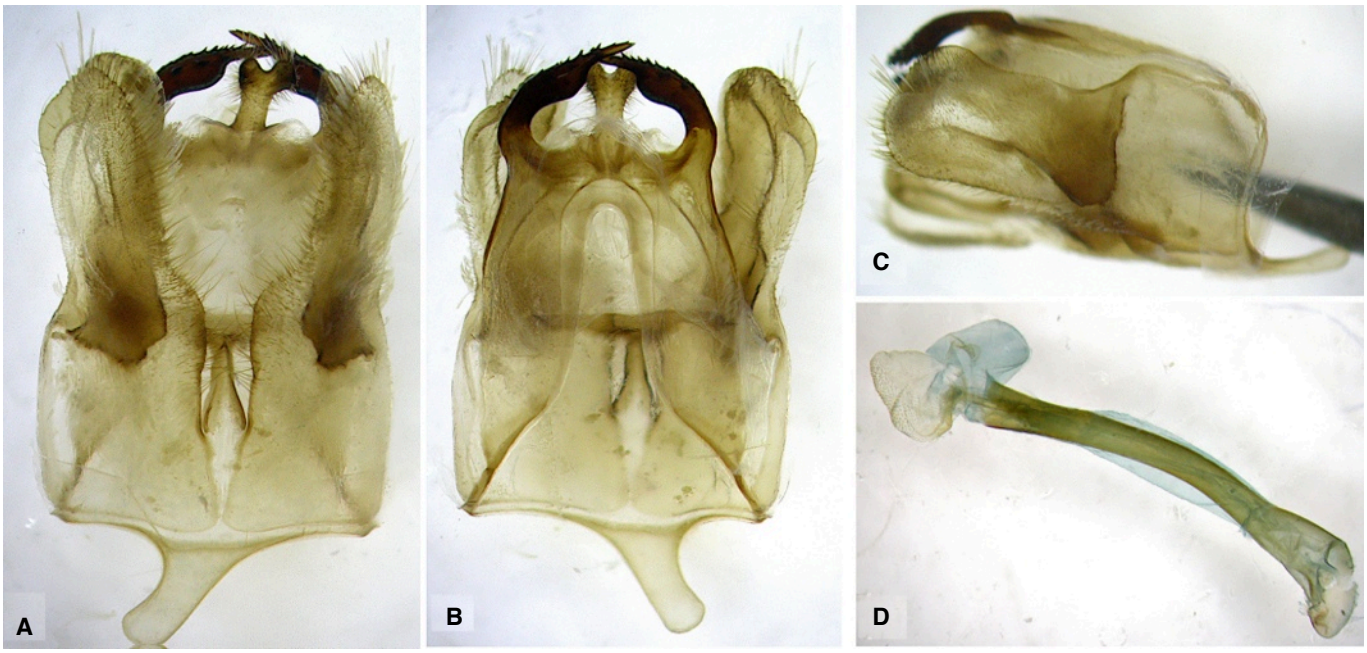

Fig. **20**. Genitalia ♂ of *Trichromia croceicervix* **sp**. **n**. from Panama (ML3855). **A**. Ventral view. **B**. Dorsal view. **C**. Lateral view. **D**. Aedeagus with everted vesica.

reach the tip of uncus. They are subquadrate at base then narrowed and subrectangular. The tip is evenly curved. Juxta short as a narrow triangle. Vinculum flat with a blade like saccus with a rounded tip and bent toward the left in ventral view. Aedeagus long, narrow and cylindrical with a short caecum penis. Vesica, short, multilobed, slightly scobinate.

Etymology. – From *croceus* which means yellow-orange and *cervix* which means here vertex, by reference to the colour of the vertex.

Early stages. – Unknown.

Distribution. – Presently known from Nicaragua to Eastern Panama.

New data and insights on various Panama Arctiinae

Very recently various forms of *Pryteria* from the Guiano-Amazonian area were synonymised under the unique name *Pryteria costata* Möschler, 1883, (Cock & Laguerre, 2021 page 259). Nevertheless, at that time, it was not possible to fix the exact status of the entity found in Central America and along the Pacific Coast due to the absence of specimen for dissection or sequencing. Fortunately in the material received, we found a pair of *Pryteria* and the male was then dissected. The genitalia from Panama and from French Guiana (see figure 21 below) are different enough to prove that the entity found in Central America and along the Pacific Coast is a full species and this confirms the name and status of this entity already predicted in Cock & Laguerre, 2021: *Pryteria colombiana* Rothschild, 1933 **bona species** which is redescribed below. The pair collected in Panama is constituted of two un-named forms: the ♂ form is close to form *tenuis* Rothschild, 1935 and the ♀ form is close to the form *apicella* (Strand, 1919) of *Pryteria costata* Möschler, 1883.

Redescription of ***Pryteria colombiana*** Rothschild, 1933

Material examined (in MLC)

- 1 ♂, PANAMA, Panama, Los Altos de Cerro Azul, 14-XI-2023, 700 m, 9.2030° N 79.4150° W, L. & J. Harrison *leg*., dissected Gen. ML3848 (ligh-blue manuscript label).

- 1 ♀, same data but 15-XI-2023.

Male (Fig. 22)

FWL = 15 mm. The male morph dissected is close to form *tenuis* described from Belize. The forewings are pure white with two black bands. One follows the internal margin from base to tornus and the second is transverse from mid-costa to tornus. This last band is noticeably wider than the same in *tenuis*. The hindwings are entirely ivory except the anal margin which is widely blackish.

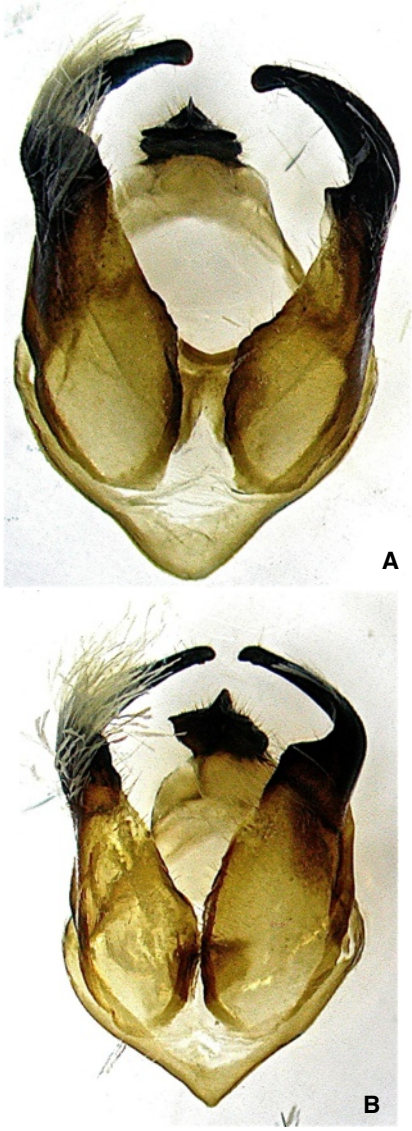

Fig. **21**. Comparison at the same scale between the genitalia ♂ of *Pryteria* from Panama (**A**) and from French Guiana (**B**).

Fig. **22**. ♂ morph of *Pryteria colombiana* from Panama, left upperside, right underside.

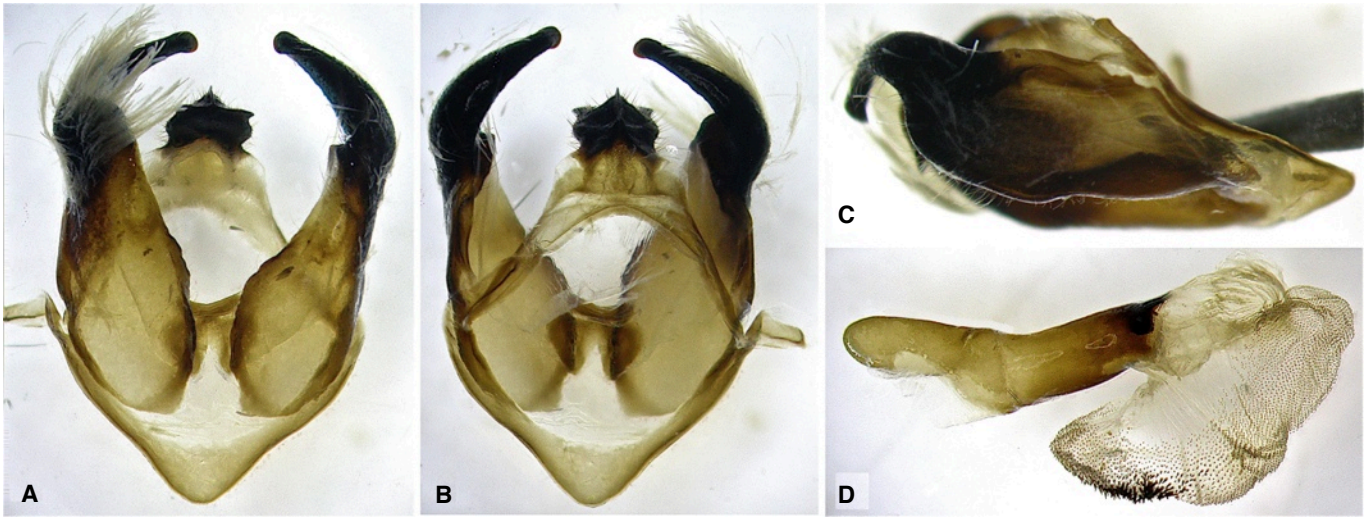

Fig. **23**. Genitalia ♂ of *Pryteria colombiana* from Panama (ML3848). **A**. Ventral view. **B**. Dorsal view. **C**. Lateral view. **D**. Aedeagus with everted vesica.

Fig. **24**. One ♀ morph of *Pryteria colombiana* from Panama, left upperside, right underside.

Genitalia male (Fig. 23)

Genital capsule slightly asymmetrical, robust and well sclerotized. Uncus strongly sclerotized flattened with a longitudinal carina. Valvae strongly sclerotized in their terminal half, clearly outreaching the tip of uncus. The right one is narrower than the left one and slightly shorter. The two are strongly bent inward and narrowed in their terminal half. Their blunt and rounded tips are in a close vicinity above uncus. Juxta concave in middle. Vinculum evenly rounded with a rounded triangular saccus. *In P. costata* the uncus is shorter and shaped as a straight and pointed triangle. Aedeagus short, straight and cylindrical with a prominent caecum penis. Vesica unilobed, bent at 180° and covered by spicules and tiny cornuti are present near the extremity.

Female (Fig. 24)

FWL = 17 mm. Morph close to morph *apicella* of *Pryteria costata*. The only difference is the total absence of blackish streaks in the apical area of forewings. The yellow lateral markings on abdomen are almost continuous whereas they are fragmented in *apicella*.

The case of *Sphecosoma stratiotes* (Dyar, 1914) **comb**. **nov**.

Dyar described *Pheia stratiotes* in 1914 on a single specimen from Trinidad River, Canal Zone, Panama which displays yellow vertex and basal forewings spots and in 1920 Schaus described *Sphecosoma roseipuncta* from a specimen from Cayuga, Guatemala with orange vertex and basal forewings spots. In his description he said "very much like stratiotes." I have sequenced two specimens: one ♀ with yellow spots from Guatemala and one ♂ with orange spots from Western Ecuador and they align at less than 0.5 %, so *roseipuncta* is merely a synonym of *stratiotes* (Fig. 25). A second specimen from Western Ecuador with orange spots confirms the identity of the sequences. Nevertheless in BOLD the two align close to *Sphecosoma* from French Guiana and not at all with *Pheia* species. Therefore I shall follow Schaus and choose *Sphecosoma* for the genus and we will have *Sphecosoma stratiotes* (Dyar, 1914) **comb**. **nov**. = *Sphecosoma roseipuncta* Schaus, 1920 **syn**. **nov**.

Material examined

- 1 ♂, PANAMA, Panama, Los Altos de Cerro Azul, 24-X-2021, 700 m, 9.2030° N 79.4150° W, L. & J. Harrison *leg*., FWL = 10 mm.

- 1 ♀, GUATEMALA, Izabal, Finca Firmeza, 18-V-2007, 680 m, 15°22.894' N 88°43.039' W, M. Laguerre *leg*., sequenced Process ID ARCTC851-11, BOLD ID MILA 1313 (yellow printed label). In MLC.

- 1 ♂, ECUADOR, Esmeraldas, 8.7 km NW Lita, 25-X-2021, 707 m, V. Doroschkin *leg*., sequenced Process ID MIAPD1779-22, BOLD ID BcHer5944. In VDC.

- 1 ♂, ECUADOR, Esmeraldas, Canandé Reserve, 30-X-2021, 314 m, 0.526675° N 79.20475° W, G. Brehm *leg*., sequenced Process ID NUTRI4225-22, BOLD ID EcEs-0140. In PMJ.

The case of the genus *Marecidia* Schaus, 1901

The genus *Marecidia* was described briefly by SCHAUS in 1901 to integrate his new species *Marecidia sanguipuncta* collected in Santa Catarina State in South-East Brazil. In 1939, FORBES, in his study of Barro Colorado Island in the Canal Zone, Panama, elaborated the description by SCHAUS and added a second species, *Marecidia achrysa*. These two species appear to be rather uncommon and almost nothing is known of their bionomics. Moreover, since their description no other specimen has been reported. The genus is well characterized in male by the dilated hind tibiae associated with the presence of an androconial pouch. Fortunately enough, I have received very recently some specimens of these poorly known species from western Ecuador and from Panama. It then appears that the locality of *M. sanguipuncta* is very likely doubtful. No specimen has been reported from S.E. Brazil but we have seen some specimens collected in Ecuador by V. Doroschkin. Moreover I have seen two similar specimens collected by G. Brehm's team in Western Ecuador, in Esmeraldas, on the road down from Lita to the Pacific Coast. These specimens are identical to the Schaus' original description but some species described by this author, allegedly from Rio de Janeiro, have in fact wrong localities. For instance *Chrysochloe sylva* (Schaus, 1896) described from Rio has never been collected in this area, but the species is rather common in the Andes Cordillera between 1500 and 2500 m. It is very likely the same situation for *Marecidia sanguipuncta* which has been collected in Western Ecuador, North and West of Lita.

On Fig 26, *M. achrysa* from Panama differs essentially from *M. sanguipuncta* from western Ecuador by the larger size of the red spot

Fig. **25**. Left *Pheia stratiotes* ♀ from Guatemala, middle *Pheia stratiotes* ♂ from Panama, right *Sphecosoma roseipuncta* ♂ from Western Ecuador.

Fig. **26**. Left ♀ *Marecidia sanguipuncta*, Western Ecuador, middle ♂ *Marecidia achrysa* Panama, right living picture of *Marecidia achrysa* in Panama (L. & J. Harrison courtesy) (not to scale).

at the forewing basis. This is clearly notified in the original description of Forbes. A specimen of *M. sanguipuncta* from Western Ecuador was sequenced and its sequence aligns at 4-4.2 % from several specimens from Peru of the recently described *Marecidia oedemeriformis* Laguerre, 2022 (BIN BOLD:AAG6243). Unfortunately we have not seen males of *M. sanguipuncta* from western Ecuador, and up to now we are unable to affirm if *achrysa* and *sanguipuncta* are conspecific or not and so we maintain the status quo.

Material examined

- **1** ♂, PANAMA, Panama, Los Altos de Cerro Azul, 29-I-2022, 700 m, 9.2030° N 79.4150° W, L. & J. Harrison *leg*., dissected Gen. ML3958 (light-blue manuscript label), FWL = 14 mm.
- **1** ♀, same data but 2-II-2022, FWL = 15 mm.
- **1** ♀, ECUADOR, Esmeraldas, 17 km N Lita, 20-XI-2021, 651 m, V. Doroschkin *leg*. In MLC.
- **1** ♀, ECUADOR, Esmeraldas, 17 km N Lita, 20-XI-2021, 651 m, V. Doroschkin *leg*., sequenced Process ID MIAPD1773-22, BOLD ID Bc-Her5938. In VDC.
- **1** ♂, ECUADOR, Esmeraldas, Canandé Reserve, 24-IV-2022, 255 m, 0.552253° N 79.1375° W, U. Diniz & D. Bottger *leg*.
- **1** ♀, same locality, but 25-X-2024, 243 m, 0.543056° N 79.1533° W, D. Rabl *leg*. Both in PMJ.

Genitalia male (Fig. 27)

Genital capsule complex. Uncus narrow and evenly bent downwards with a pointed tip. On the tegumen on each side of the uncus a rounded shoulder. Valvae divided in two parts. Ventrally an elongated and translucent lobe covered with thin hairs reaching almost the tip of uncus. Dorsally a narrow and shorter process, bifid at the extremity and narrowed at two thirds. An arched, narrow and long process starting from the base of valvae and largely outreaching the tip of uncus. This process ends with a lanceolate and pointed extremity, denticulate on side. Vinculum convex with a short pointed triangular saccus. Aedeagus short, cylindrical, slightly bent with a single lobed vesica covered with spicules.

The case of *Rhipha persimilis marginata* (Rothschild, 1909)

In 1909, Rothschild described *Automolis persimilis* from Peru, La Oroya, R. Inambari (lectotype ♂ in NHMUK) and in the same paper he described the ssp *marginata* from Costa Rica, Tuis (lectotype ♂ in NHMUK). In 1975 Watson transferred these two entities within the genus *Rhipha* Walker, 1854 (Watson, 1975 p. 91). In the Catalogue published in 2014 these two entities were considered as synonyms. One male specimen was found in the material received from Panama (see ♂ below Fig. 28 and ♂ genitalia Fig. 29). The genitalia are identical to the dissection of the lectotype of *marginata* provided by A. Watson in NHMUK (reference 1686). Unfortunately the genitalia are very homogeneous in this group and it is not possible to confirm that *marginata* is a synonym or a subspecies of *persimilis* and not a full species. Only sequencing will be able to solve this problem.

Fig. **27**. Genitalia ♂ of *Marecidia achrysa* from Panama (Gen. ML3958). **A**. Ventral view. **B**. Dorsal view. **C**. Lateral view. **D**. Aedeagus with everted vesica.

Fig. **28**. ♂ of *Rhipha persimilis marginata* from Panama, left upperside, right underside, label not to scale.

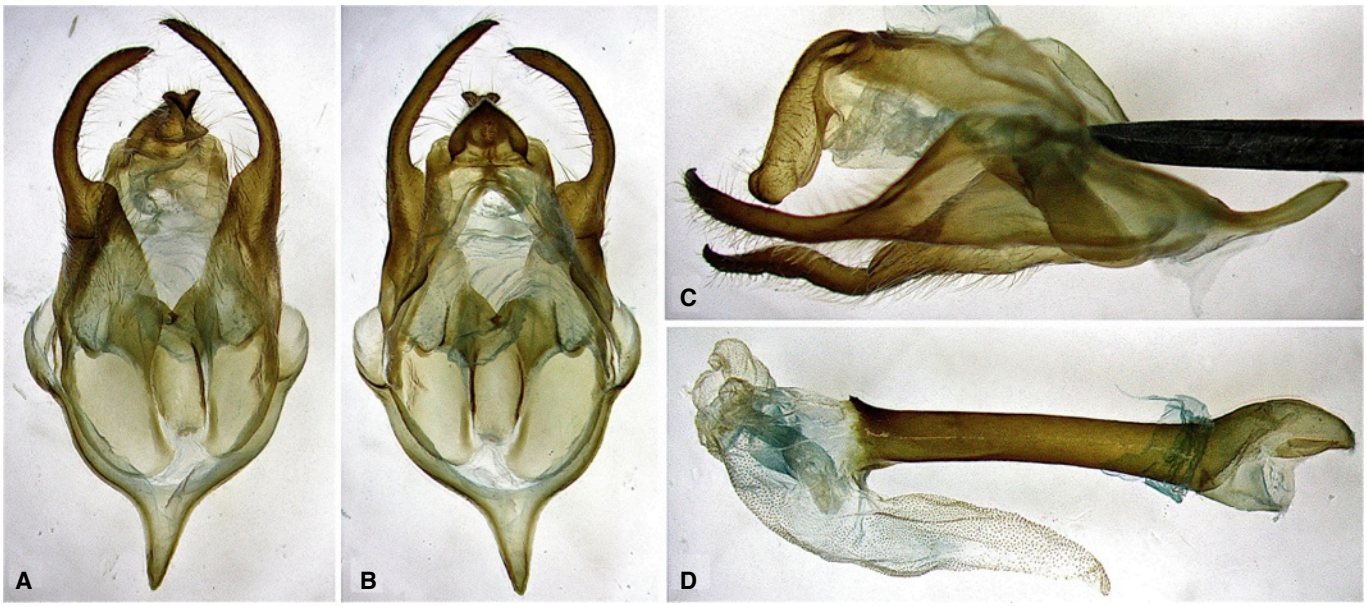

Fig. **29**. Genitalia ♂ of *Rhipha persimilis marginata* from Panama (ML3880). **A**. Ventral view. **B**. Dorsal view. **C**. Lateral view. **D**. Aedeagus with everted vesica.

Fig. **30**. ♀ of unknown *Regobarrosia sp.* from Panama, left upperside.

Material examined

- 1 ♂. PANAMA, Panama, Los Altos de Cerro Azul, 05-VIII-2023, 700 m, 9.2030° N 79.4150° W, L. & J. Harrison *leg*., dissected Gen. ML3958 (light-blue manuscript label). In MLC. FWL = 14 mm.

Genitalia male (Fig. 29)

Genital capsule slightly assymetrical not unlike the male genitalia of *Pseudalus*: right valva slightly longer and less curved than left valva. Uncus bent at 90° downward with two dorsal carinae fused at tip. Tegumen flat on each side of the uncus. Valvae large at base then strongly narrowed in the last half, finishing as a narrow and pointed arm. The right one is longer and weakly bent inward, the left one is shorter and more strongly bent inward. Juxta ovoid. Vinculum evenly rounded with a long and narrow pointed saccus. Aedeagus straight, narrow and cylindrical. Caecum penis present. Vesica with a single lobe shaped as a tongue turned at 180° and covered with tiny spicules.

The case of the genus ***Regobarrosia*** Watson, 1975

Within the material received, a single *Regobarrosia* was found. In fact no species of this genus has been reported from Central America and this specimen represents maybe a new species (see Fig. 30). Unfortunately it is a female and, in the absence of a sequence or subsequent male genitalia, it is impossible to establish its exact status.

Material examined

- 1 ♀. PANAMA, Panama, Los Altos de Cerro Azul, 19-III-2023, 700 m, 9.2030° N 79.4150° W, L. & J. Harrison *leg*., in MLC. FWL = 15 mm.

The case of ***Hyperthaema haemacta*** Schaus, 1901

Among the material received from Panama, there was a male specimen of the genus *Hyperthaema* Schaus, 1901 (see Fig. 31). This specimen resembles *H. haemacta* Schaus, 1901 described from Costa Rica, Cartago, El Sitio (lectotype in USNM). The specimen was dissected and its genitalia were compared to the lectotype dissection made by Alan Watson (Watson, 1971, Pl. 156, reference AW255) thus confirming the identity of the specimen. A picture of the fresh specimen along with a description of its genitalia are provided below (Fig. 32).

Material examined

- 1 ♂, PANAMA, Panama, Los Altos de Cerro Azul, 11-XI-2023, 700 m, 9.2030° N 79.4150° W, D. Herbin *leg*., dissected Gen. ML3831 (light-blue manuscript label). In MLC. FWL = 16 mm.

Genitalia male (Fig. 32)

Genital capsule assymetrical, right valva longer than left valva. Uncus short, well sclerotized bent downward with a blunt tip. Below uncus a subquadrate structure (maybe two fused socii). The valvae are long, outreaching the tip of uncus. Their base is large, then they are divided into two parts: one ventral narrow and elongated and one dorsal spatulated at tip. The right valva is longer than the left one, the ventral part is narrow and slightly bent dorsally, longer than the dorsal part which is straight and spatulated at tip. For the left valva, the two parts are subequal and shorter than the right ones. The ventral one is narrower and straight, the dorsal one is larger, straight and spatulated at tip. Juxta subrectangular divided vertically by a cleft. Vinculum and saccus shaped as a pointed triangle slightly bent to the left in ventral

Fig. **31**. ♂ of *Hyperthaema haemacta* from Panama, left upperside, right underside.

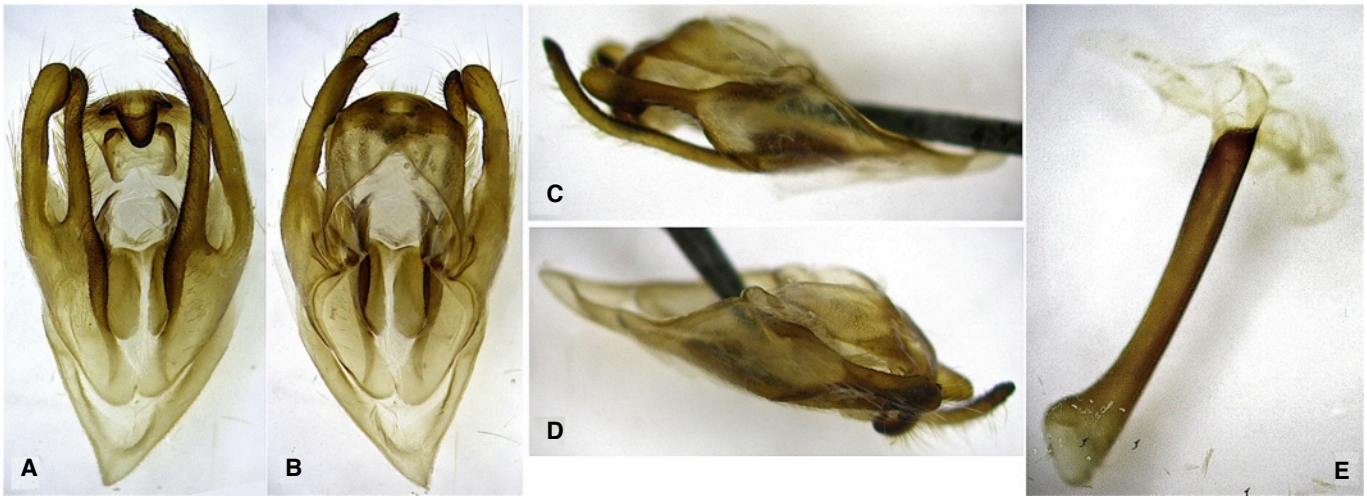

Fig. **32**. genitalia ♂ of *Hyperthaema haemacta* from Panama (ML3831). **A**. Ventral view. **B**. Dorsal view. **C**. Right lateral view. **D**. Left lateral view. **E**. Aedeagus with everted vesica.

view. Aedeagus straight, narrow and cylindrical. Vesica smooth, perpendicular to the aedeagus with one lobe on each side.

Below is included a list of some interesting material collected in the same area in Panama. The list is presently far from being exhaustive and more interesting species will very likely be discovered with time.

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Annex: list of some interesting material collected in Panama

The data are classified alphabetically within their various subtribes. The nomenclature used is the present one even if in a lot of cases this nomenclature is largely obsolete.

ARCTIINI Leach**,** 1815

PHAEGOPTERINA Kirby, 1892

Amaxia pandama (Druce, 1893)
Arctiarpia melanopasta (Dognin, 1907)
Castrica phalaenoides (Drury, 1773)
Cissura plumbea Hampson, 1901
Coiffaitarctia vincentii Gibeaux, 2020
Elysius diorus Laguerre, 2024
Elysius rubromarginatus Laguerre, 2024
Emurena tripunctata (Druce, 1884)
Evius cochenouri Schaus, 1910
Glaucostola romula (Druce, 1895)
Haemanota haemabasis (Dognin, 1914)
Haemaphlebiella chrysophlebia Laguerre, 2021
Hyperandra excavata (Forbes, 1939)
Idalus dares Druce, 1894
Idalus carinosa (Schaus, 1905)
Idalus aleteria (Schaus, 1905)
Idalus perlineosa (Rothschild, 1917)
Idalus septentrionalis Laguerre, 2016
Ischnognatha leucapera (Dognin, 1914)
Kirrostola metaxantha (Schaus, 1905)
Melese asana (Druce, 1884)
Melese flavimaculata Dognin, 1899
Metaxanthia atribasis Rothschild, 1913
Neonerita bernardoespinozai Laguerre & Vincent, 2016
Ochrodota pronapides (Druce, 1894)
Parathyris griseata (Rothschild, 1935)
Phaeomolis lineatus (Druce, 1884)
Psychophasma erosa (Herrich-Schäffer, [1858])
Rhipha chionoplaga (Dognin, 1913)
Rhodorhipha cf. flammans (Hampson, 1901)
Robinsonia bartolana Laguerre & Espinoza, 2006
Robinsonia deiopea Druce, 1895
Selenarctia schausi (Rothschild, 1916)
Symphlebia hyalina amaculata (Rothschild, 1910)
Symphlebia ipsea (Druce, 1884)
Symphlebia janecheverriae Espinoza, 2017
Symphlebia perflua rosea (Schaus, 1894)
Thysanoprymna hampsoni (Dognin, 1902)
Trichromia atta (Schaus, 1920)
Trichromia cardinalis (Dognin, 1899)
Trichromia cotes (Druce, 1896)
Tricypha imperialis (Heylaerts, 1884)
Viviennea tegyra (Druce, 1896)
Watsonidia reimona (Schaus, 1933)

SPILOSOMINA Kirby, 1892

Oxyptera magdalenae (Oberthür, 1881)

PERICOPINA Walker, 1869

Chetone zuleika Becker & Goodger, 2013
Dysschema viuda (Schaus, 1910)
*Dysschema jansonis (*Butler, [1870])
Eucyane arcaei Druce, 1884
Eucyane reedia (Schaus, 1910)
Hyalurga fenestra (Linnaeus, 1758)
Hyalurga sixola Schaus, 1910
Hyalurga urioides Schaus, 1910
Isostola philomela (Druce, 1893)
X*enosoma geometrina* (Schaus, 1910)

EUCHROMIINA Butler, 1876

Autochloris jansonis (Butler, 1872)
Calonotos tiburtus (Cramer, [1779])
Cosmosoma angustimargo Schaus, 1911
Cosmosoma regia (Schaus, 1894)
Cosmosoma remotum (Walker, 1854)
Cosmosoma semifulva (Druce, 1884)
Cosmosoma stilbosticta (Butler, 1876)
Cosmosoma zurcheri Druce, 1894
Gymnelia colona (Schaus, 1911)
Hypocharis clusia (Druce, 1897)
Lepidoneiva caecum (Hampson, 1898)
Loxophlebia flavipicta Schaus, 1912
Phoenicoprocta paucipuncta Dyar, 1914
Phoenicoprocta sanguinea (Walker, 1854)
*Planusuncus hercyna (*Druce, 1884)
Planusuncus hercynacula (Dyar, 1915)
Psoloptera thoracica (Walker, 1854)
Saurita anselma Schaus, 1924
Saurita rubripuncta Schaus, 1911

Saurita sanguinea (Druce, 1884)
Saurita watsoni Rotschild, 1911
Xanthyda chalcosticta (Zerny, 1931)
Xanthyda saron (Druce, 1884)
Xanthyda xanthosticta (Hampson, 1898)

CTENUCHINA Kirby, 1837

Aclytia albistriga Schaus, 1910
Acridopsis audureaui Cerda, 2020
Aderithales atrigutta (Druce, 1905)
Antichloris viridis Druce, 1884
Anycles anthracina (Walker, 1854)
Argyroeides notha Schaus, 1911
Cercopimorpha sylva Schaus, 1920
Coccabdominis balium (Hampson, 1898)
Delphyre rhodocrypta (Druce, 1905)
Ecdemus obscuratum Schaus, 1911
Epanycles imperialis (Walker, 1854)
Episcepsis aelia (Schaus, 1889)
Episcepsis hypoleuca Hampson, 1898
Eriphioides tractipennis (Butler, 1876)
Eucereon consorta (Schaus, 1910)
Eucereon dognini Rothschild, 1912
Eucereon facundum Draudt, 1917
Eucereon venosa Schaus, 1910
Flammeus walkeri (Druce, 1889)
Gloora alba (Druce, 1894)
Heliura solicauda Butler, 1876
Heliura thysbodes Dognin, 1914
Hermierius testaceus (Druce, 1884)
Horama plumipes (Drury, 1773)
Hypocladia restricta Hampson, 1901
Laguerrellus banoca (Dyar, 1914)
Leucopleura ciarana Schaus, 1924
Nelphe relegatum (Schaus, 1911)
Pseudosphex moza (Druce, 1896)
Pupillaptera completa (Draudt, 1916)
Telioneura obsoleta (Draudt, 1915)
Theages decora (Schaus, 1910)
Theagess flavicaput (Hampson, 1898)
Theages xanthura (Schaus, 1910)
Uranophora leucotelus (Butler, 1876)

LITHOSIINI Billberg, 1820

Apistosia judas Hübner, 1827

Résumé

Laguerre M., 2025. – Nouvelles espèces d'Arctiinae à l'Est de la Zone du Canal de Panama (Lepidoptera, Erebidae, Arctiinae, Arctiini). *Faunitaxys*, 13(12): 1 – 17.

L'étude d'un matériel récent récolté au Panama à l'Est de la zone du Canal a révélé cinq nouvelles espèces qui sont décrites : *Robinsonia xanthopoda* **sp**. **n**., *Syntomostola harrisoni* **sp**. **n**., *Pseudosphex lindae* **sp**. **n**., *Pseudalus panamensis* **sp**. **n**., et *Trichromia croceicervix* **sp**. **n**. De plus de nouveaux détails pour des espèces peu connues ont permis de nouvelles recombinaisons, synonymies ou l'établissement de nouveaux status pour plusieurs espèces dans les genres *Pryteria* Möschler, 1883, *Marecidia* Schaus, 1901 ou *Sphecosoma* Butler, 1876. Enfin les genitalia mâles sont illustrés pour la première fois pour quelques espèces dans les genres *Rhipha* Walker, 1854 ou *Hyperthaema* Schaus, 1901.

Mots-clés. – Phaegopterina, Euchromiina, Ctenuchina, taxonomie, nouvelles espèces, description, Canal de Panama.

Derniers articles publiés

Lourenço W. R. & Ythier E., 2025. – A new cave species of *Isometrus* Ehrenberg 1828 from Myanmar (Scorpiones: Buthidae). *Faunitaxys*, 13(04): 1 – 6. https://doi.org/10.57800/faunitaxys-13(04)

Platia G. & Pedroni G., 2025. – Five new species and new records of click beetles from Saudi Arabia with an updated catalogue (Coleoptera, Elateridae). *Faunitaxys*, 13(05): 1 – 12. https://doi.org/10.57800/faunitaxys-13(05)

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Lourenço W. R. & Velten J., 2025. – Description of a second species for the genus *Serratochaerilobuthus* Lourenço, 2024 (Scorpiones: Chaerilobuthidae). *Faunitaxys*, 13(10): 1 – 6. https://doi.org/10.57800/faunitaxys-13(10)

Jarzabek-Müller A., 2025. – A new *Idolus* Desbrochers des Loges, 1875 for the Caucasus region (Coleoptera, Elateridae, Pomachiliini). *Faunitaxys*, 13(11): 1 – 6. https://doi.org/10.57800/faunitaxys-13(11)

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SOMMAIRE

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Illustration de la couverture :

Living picture of Hyalurga sixola Schaus, 1910, male on Cerro Azul in Eastern Panama (credit photo: L. & J. Harrison).

Crédits:

M. **Laguerre** : Fig. 2-5, 7-9, 11-25 & 27-32.
L. & **J**. **Harrison** : Fig. 1, 6, 10, 26 & couverture.

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